

“OPTIMIZATION OF GROWTH CONDITION FOR ACIDIC PECTINASE AT SUBMERGED FERMENTATION FROM FRUIT WASTE.”

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ABSTRACT

Three fungus strains were isolated from compost to make Pectinase. It was determined that cultivation of Talaromyces aurantiacus, Penicillium oxalate and Aspergillus niger on orange peel substrate under various SSF conditions is a low-cost alternative to submerged-liquid fermentation when it comes to pectinase production. Optimization of environmental conditions for Pectinase production was carried out using Pectinase screening agar media (PSAM). Among other factors, the parameters that concern temperature and pH. The findings show the rise in temperature 25°C and pH 4 shows decent growth. The greatest harvest occurred at orange peels, which yielded 28mm enzymatic properties and yield might be improved by PSAM optimization.

KEYWORDS: Pectinases, Enzyme, Optimization, Pectinase, Solid State Fermentation

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INTRODUCTION

The processing of citrus fruits generates over 34 million tonnes of citrus residue (peel, seeds) per year [Huang et.al., (2010), Martín et.al., (2010), Anwar et.al., (2008)]. Many environmental issues are caused by a small fraction of these residues being applied in the diet of monogastric animals. While citrus peel is an essential and affordable raw material for producing enzymes, most notably Pectinase, via fermentation, it may also be used to prepare citrus oil, which can then be used for a variety of purposes [Palit et.al., (2001)]. The media that is used in this experiment provides all of the nutrients required for microorganism development, and numerous key metabolites, including enzymes, have been examined in regard to their synthesis. Extraction, clarity, and cloud stabilization of fruit juices, in degumming and retting of natural fibers are all assisted by Pectinases, which catalyze the breakdown of pectin (polygalacturonic acid) to galacturonic acid residues [Kumar et.al., (2010) Mamma et.al., (2008) Tari et.al., (2007), Botella et.al., (2007), Kuhad et.al., (2004), De Gregorio et.al., (2002), Silva et.al., (2002)].

Using pectinolytic enzymes, different industrial processes may be handled. Because these enzymes are effective at decreasing the viscosity of liquors during the clarifying process, they are employed widely in the food and drink industries, mainly in juice clarity [Oszmianski et.al., (2011), Pedrolli et.al., (2008)]. However, prior studies show that PG is produced by Solid State Fermentation using agro-industrial wastes and other microorganisms. Consider the metabolites generated in an Solid State Fermentation to be trapped in a solid matrix, requiring them to be removed via a liquid-liquid extraction or leaching. Since this is the first stage in every recovery and purification procedure for any desired metabolite created by Solid State Fermentation, this is also the first step in any process like that.

Due to their low cost, significant enzyme production capacity, and affordable cost, orange peel (pectin waste) is utilized as a substrate rather as synthetic cellulose, resulting in reduced manufacturing costs [Sharma et.al.,

2004]. The reduction in cost makes the Pectinase manufacturing process much more economically feasible. A serious problem has to be solved. As in the case of fermentation, which harnesses pectin enzymes to help manufacturers reduce waste and pollution, 2 of 9 with the residue for complete use of pectin resources and to keep pollutants out of the environment [He et.al, (2004)].

Varying the only independent variable while maintaining the others at a fixed level is the typical technique for improving enzyme production by utilising a “one variable at a time” strategy. By utilising this method, it is tedious and time-consuming as it necessitates the use of a huge number of variables, and it does not factor in how various variables interact [Kumari et.al., (2008)]. Statistics can be used as an alternate and more efficient technique. In PSAM, a thorough factorial search of all the factors is made in order to find all of the simultaneous, systematic, and efficient variation. While statistical optimization makes it possible to quickly and thoroughly test a whole large-scale experiment, it also reflects the contributions of each of the components. The implementation of PSAM for media and culture conditions has previously proved to be effective when used to maximise productivity in the synthesis of primary and secondary metabolites, such as amino acids, ethanol, and enzymes.

This research was conducted to improve the production of an acidic pectinase from Fungal spp., as well as to explore industrial possibilities for the enzyme, in light of the newly-found fact that pectinases has industrial uses. Using Central Composite Design and PSAM, the factors that were shown to be essential to optimising enzyme production in submerged fermentation were optimised, and hence, increased.

MATERIALS AND METHODS

Substrates and Chemicals

Orange peels taken from the market in Satna, Madhya Pradesh, India were found to contain a variety of agro-waste residues. A supply of fine-ground substrates was dried in the hot air oven at 60 degrees Celsius (C) for 48 hours, ground, and stored in sterile containers until use. This study used all Hi-Media Limited, SRL Pvt. Limited-made chemicals.

Source of Fungal Inoculums

A fungus species that produces pectinase was isolated from Malt extract agar medium (which contains plenty of pectin) and cultured at 4 °C.

Solid State Fermentation

Dried citrus peel (which had a 75% average particle size ranging from 0.8 to 2.0 mm, and the other 25% had a range between 2.0 and 3.0 mm) was mixed with the following substances, as a dry substance, which supplies the following elements: Ammonium nitrate (0.43%); sodium sulphate (0.021%); magnesium sulphate (7.7H₂O, 0.077%); zinc sulphate (0.042%); potassium chloride (0.162%); and calcium hydroxide (0.011%). Adding water to increase the percentage of initial humidity got us to 60 percent. One part inoculant was used in a tenfold (1:10) dilution of moist solid medium. The flask was 250 mL in size and the neck was broad, and therefore was appropriate for fermentations. Cultures were left to develop at their own pace without being agitated. Every experiment was performed three times.

Polygalacturonase (PG) activity

The D-N-Salicylsalicylic (DNS) assay was used to determine the level of reducing sugars present in the test sample. Pectin was a simple sugar produced following pectin breakdown, and galacturonic acid (Sigma) was regarded as a simple sugar to

that particular study[Breed et.al., (1947)]. When diluted, an enzyme solution had 0.1 mL of the enzyme solution added to 0.9 mL of 0.5% (w/v) pectin (Sigma) in 0.1 M citrate buffer (pH 4.0). The reaction was allowed to incubate at 50 °C for 15 minutes, after which it was heated to boiling water for 5 minutes. To each sample, a total of 5 mL of distilled water was added. Samples were examined for absorbance at 540 nm. The activity of a single enzyme that catalyses the conversion of D-galacturonic acid to D-galactose is defined as 1 mol of D-galacturonic acid released per minute at 50°C and pH 4.0 [Rodríguez-Fernández et.al., (2011)].

Optimization Design

Various microbial strains, media, and growth factors were systematically and individually varied. 3 strains had the ability to degrade pectin *Talaromyces aurantiacus*, *Penicillium oxalate*, and *Aspergillus niger*, all performed within a temperature range of 20 to 35 degrees Celsius with a pH of 2 to 5 at substrate such orange peel, banana peel, sugarcane bagasse, and pineapple peel.

Result and Discussion

Three strains of fungus were selected following the first screening, where each was shown to have pectinase activity. This demonstrates the outcomes of the studies on pH and temperature optimization, as well as variable substrate optimization.

Temperature Optimization Results

Graph for Temperature optimization and shows that Acidic Pectinase enzyme activity is highest at 25°C.

Table 1: Culture Optimization and Enzyme Characterization (Temperature)

| S.No. | Temperature(°C) | Growth (mm in 7 days) |
|-------|------------------|-----------------------|
| 1 | 20 | 20 |
| 2 | 25 | 28 |
| 3 | 30 | 26 |
| 4 | 35 | 11 |

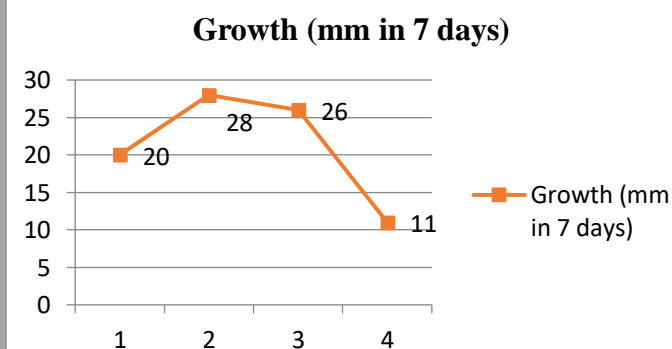


Table 2: Culture Optimization and Enzyme Characterization (pH)

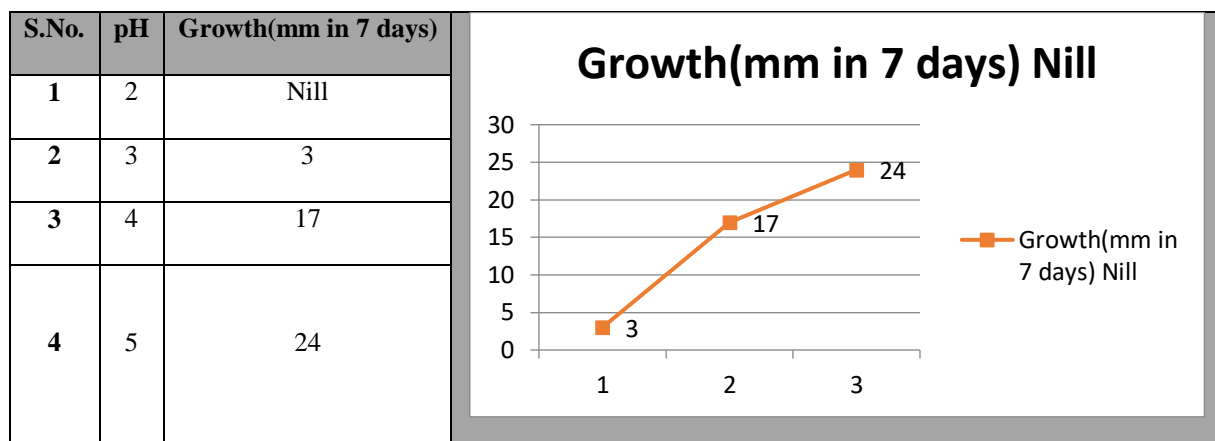


Table 3: Culture Optimization and Enzyme Characterization (Substrate)

| S.No. | Nutrients(Fruit Waste) | Growth (mm in 7 days) |
|-------|------------------------|-----------------------|
| 1 | Orange peel | 20 |
| 2 | Sugarcane baggases | 8 |
| 3 | Banana peel | 3 |
| 4 | Apple peel | Nil |

CONCLUSIONS

The research was done to improve enzyme manufacturing conditions. As a result, costs for Acidic Pectinase are increasing, resulting in increased fruit juice clearance and a wider range of industrial uses. Thus, as a result, the fact that Pectinase increases in production at pH 4, 25°C, and with orange peels.

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